

## RESEARCH LETTER – Pathogens &amp; Pathogenicity

# AsaGEI2b: a new variant of a genomic island identified in the *Aeromonas salmonicida* subsp. *salmonicida* JF3224 strain isolated from a wild fish in Switzerland

Jean-Guillaume Emond-Rheault<sup>1,2,3</sup>, Antony T. Vincent<sup>1,2,3</sup>, Mélanie V. Trudel<sup>1,2,3</sup>, Joachim Frey<sup>4</sup>, Michel Frenette<sup>3,5</sup> and Steve J. Charette<sup>1,2,3,\*</sup>

<sup>1</sup>Institut de biologie intégrative et des systèmes, Pavillon Charles-Eugène-Marchand, Université Laval, 1030 avenue de la Médecine, Quebec City, QC G1V 0A6, Canada, <sup>2</sup>Centre de recherche de l'Institut universitaire de cardiologie et de pneumologie de Québec (Hôpital Laval), 2725 Chemin Sainte-Foy, Quebec City, QC G1V 4G5, Canada, <sup>3</sup>Département de biochimie, de microbiologie et de bio-informatique, Faculté des sciences et de génie, Université Laval, 1045 avenue de la Médecine, Quebec City, QC G1V 0A6, Canada, <sup>4</sup>Institut of Veterinary Bacteriology, University of Bern, Länggassstrasse 122, CH-3012 Bern, Switzerland and <sup>5</sup>Groupe de Recherche en Écologie Buccale (GREB), Faculté de médecine dentaire, Université Laval, Quebec City, QC G1V 0A6, Canada

\*Corresponding author: Institut de Biologie Intégrative et des Systèmes (IBIS), Pavillon Charles-Eugène-Marchand, 1030 avenue de la Médecine, Université Laval, Quebec City, QC G1V 0A6, Canada. Tel: +418-656-2131, ext. 6914; Fax: +418-656-7176; E-mail: [steve.charette@bcm.ulaval.ca](mailto:steve.charette@bcm.ulaval.ca)

**One sentence summary:** Identification of AsaGEI2b, a new genomic island as an additional indicator of the geographic origin of *Aeromonas salmonicida* subsp. *salmonicida* isolates.

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## ABSTRACT

*Aeromonas salmonicida* subsp. *salmonicida* is the causal agent of furunculosis in salmonids. We recently identified a group of genomic islands (AsaGEI) in this bacterium. AsaGEI2a, one of these genomic islands, has almost exclusively been identified in isolates from North America. To date, *Aeromonas salmonicida* subsp. *salmonicida* JF3224, a strain isolated from a wild brown trout (*Salmo trutta*) caught in Switzerland, was the only European isolate that appeared to bear AsaGEI2a. We analyzed the genome of JF3224 and showed that the genomic island in JF3224 is a new variant of AsaGEI, which we have called AsaGEI2b. While AsaGEI2b shares the same integrase gene and insertion site as AsaGEI2a, it is very different in terms of many other features. Additional genomic investigations combined with PCR genotyping revealed that JF3224 is sensitive to growth at 25°C, leading to insertion sequence-dependent rearrangement of the locus on the pAsa5 plasmid that encodes a type three secretion system, which is essential for the virulence of the bacterium. The analysis of the JF3224 genome confirmed that AsaGEIs are accurate indicators of the geographic origins of *A. salmonicida* subsp. *salmonicida* isolates and is another example of the susceptibility of the pAsa5 plasmid to DNA rearrangements.

**Keywords:** AsaGEI2b; genomic island; *Aeromonas salmonicida* subsp. *salmonicida*; type three secretion system

## INTRODUCTION

The Gram-negative bacterium *Aeromonas salmonicida* subsp. *salmonicida* is an opportunistic fish pathogen that is responsible for significant economic losses in salmonid fish farms (Bernoth et al. 1997). It is the causal agent of furunculosis, a predominantly salmonid disease with high morbidity and mortality (Janda and Abbott 2010).

The international fish trade may contribute to introducing pathogens into new environments (Hedrick 1996). Additional epidemiological tools are thus required to track the dissemination of *A. salmonicida* subsp. *salmonicida* isolates into new ecosystems.

We recently described a new genomic island (*Aeromonas salmonicida* genomic island, AsaGEI) in this bacterial species (Emond-Rheault et al. 2015). Genomic islands (GEIs) are genetic elements ranging in size from 10 to 200 kb that are inserted in the bacterial chromosome. They are not present in all strains of a species but can provide various functional advantages to those possessing them, including increased pathogenicity (Juhas et al. 2009; Bellanger et al. 2013; Darmon and Leach 2014). To date, we have identified three forms of AsaGEI (1a, 1b and 2a) that range in size from 50 to 53 kb. They all contain many phage genes. Their contribution to the host phenotype is unknown (Emond-Rheault et al. 2015).

Nevertheless, AsaGEIs are interesting from a molecular epidemiological point of view. Each AsaGEI appears to be strongly associated with a specific geographic region, suggesting that they can be used as indicators of geographic origin. AsaGEI1a has only been found in the Great Lakes-St. Lawrence River system, while AsaGEI1b, except for two isolates, was found in Europe. On its side, AsaGEI2a has been exclusively detected in North American isolates, with the exception of JF3224 (Emond-Rheault et al. 2015), which was isolated from a wild brown trout (*Salmo trutta*) captured in 2004 in a prealpine Swiss river (Burr et al. 2005). More precisely, of 139 isolates tested, 82 bore an AsaGEI2a, while only one of 25 isolates from Europe (JF3224) harbored this GEI. This exception prompted us to investigate this isolate using whole genome sequencing in order to determine whether it was an example of human-driven pathogen dissemination. In fact, our analyses revealed that this isolate contains a novel variant of AsaGEI.

## Materials and Methods

### Bacterial isolates and growth conditions

The *A. salmonicida* subsp. *salmonicida* isolates used in the present study were JF3224, which was isolated from a wild brown trout (*Salmo trutta*) captured in 2004 in a prealpine Swiss river (Burr et al. 2005), and 01-B526, which was used as a reference strain with an intact pAsa5 plasmid (Daher et al. 2011; Charette et al. 2012). They were grown for 2 or 3 days at 18°C on furunculosis agar (10 g of Bacto-Tryptone, 5 g of yeast extract, 1 g of L-tyrosine, 2.5 g of NaCl and 15 g of agar per liter of distilled water) (Hanninen and Hirverikoski 1997).

### DNA extraction and genomic sequencing

The total genomic DNA of JF3224 was extracted using DNeasy blood and tissue kits (Qiagen, Canada). The TruSeq shotgun library was generated using purified genomic DNA and was sequenced by the Plateforme d'Analyse Génomique of the Institut de Biologie Intégrative et des Systèmes (IBIS, Université Laval) using Illumina technology on a MiSeq instrument.

Table 1. Primers used in this study.

Primer	Sequence (5'–3')	Reference
Control primers for pAsa5		
traC F	5'-TGCACTATCCCCAGCTATCC-3'	(Daher et al. 2011)
traC R	5'-TCGGTAATCGCGGTCTTGTC-3'	(Daher et al. 2011)
exsD F	5'-AGAAGTGATCCTGACCCAAGGCAA-3'	(Daher et al. 2011)
exsD R	5'-TTGCAACGACTGTTGCCAAGAACC-3'	(Daher et al. 2011)
resD F	5'-TCAGAAACTTGGCCATCGCTCACA-3'	(Daher et al. 2011)
resD R	5'-TGATGTGCAGATTTCCTGGAGCA-3'	(Daher et al. 2011)
B–C rearrangement primers		
11B1F	5'-GCGCACCACCACCATTTAATGTCA-3'	(Tanaka et al. 2012)
11CR	5'-AACTGGCAAGGATAGAGCTGCTGA-3'	(Tanaka et al. 2012)

### Sequence analyses

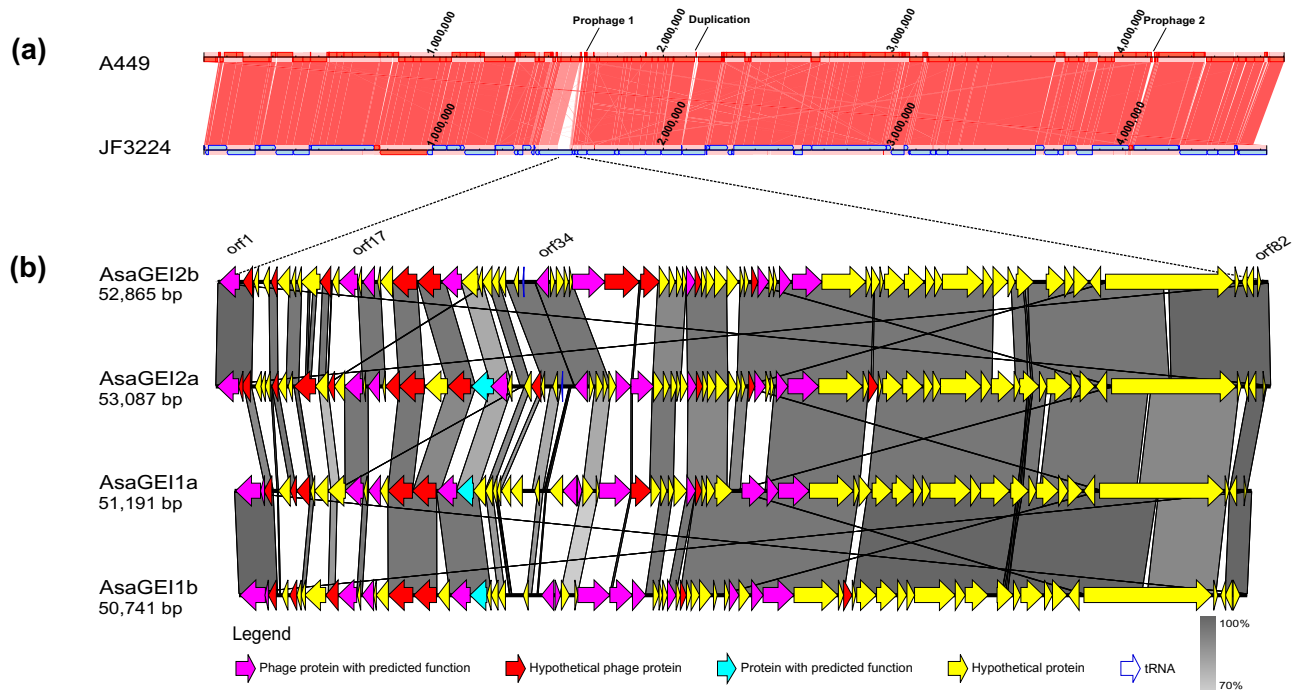
A total of 1800 692 raw reads (1702 181 after quality control and error correction) with an average length of 300 bp were generated (mean coverage: 86.53×) and were *de novo* assembled using A5-miseq pipeline version 20140401 (Coil, Jospin and Darling 2015). The complete structures of the chromosome of the JF3224 isolate and the pAsa5 plasmid were determined by genomic alignment using the CONTIGuator web-server (Galardini et al. 2011) with the chromosome (GenBank accession number: NC.009348.1) and pAsa5 plasmid (GenBank accession number: NC.009350) from the A449 strain as reference sequences (Reith et al. 2008). The Whole Genome Shotgun project of *A. salmonicida* subsp. *salmonicida* JF3224 has been deposited in the DDBJ/EMBL/GenBank database under accession number JXTA00000000. The draft genome sequences were annotated using the NCBI Prokaryotic Genome Annotation Pipeline.

AsaGEI2b was annotated as previously described (Emond-Rheault et al. 2015). Briefly, the open reading frames (ORFs) were predicted by Prodigal (version 1.20) (Hyatt et al. 2010), confirmed by GeneMark (version 2.5) (Besemer, Lomsadze and Borodovsky 2001) and visualized in Artemis (version 16.0.0) (Carver et al. 2012). Each ORF was manually verified. BLASTn and BLASTp searches (NCBI) were performed to determine the putative functions of the ORFs (Altschul et al. 1997). The non-redundant (nr/nt) and whole-genome shotgun nucleotide databases were used for the BLASTn search and the non-redundant protein sequences (nr) database was used for the BLASTp search.

### PCR analyses

The DNA templates, PCR mixtures and program cycles were as previously described (Trudel et al. 2013). The PCR assays were performed in duplicate with the appropriate positive and negative controls for each condition. The PCR primers used are listed in Table 1.

To compare the pAsa5 plasmid from the JF3224 isolate in Switzerland to the pAsa5 plasmid in the isolate shipped to Canada as well as its susceptibility to incubation at 25°C, the same PCR assays were performed in Switzerland on the original isolate grown on tryptic soy agar for 3 days at 18°C or 25°C. A pool of approximately 20 colonies grown at 18°C or 25°C were suspended in 450 µl of lysis buffer (100 mM Tris-HCl, pH 8.5, 0.05% Tween 20 and 240 µg ml<sup>-1</sup> of proteinase K) and were incubated for 1 h at 60°C and then for 15 min at 97°C. The cell lysates were used as DNA templates for the PCR assays described above.



**Figure 1.** Identification of AsaGEI2b in the genome of the JF3224 *A. salmonicida* subsp. *salmonicida* isolate. (a) Chromosomal alignment of the JF3224 contigs with the A449 reference strain (Reith et al. 2008) using the CONTIGuator webserver (Galardini et al. 2011). (b) Representation of the genomic alignments of the new AsaGEI element in the genome of JF3224 (AsaGEI2b) with the other three AsaGEIs identified in a previous study (Emond-Rheault et al. 2015) using Easyfig (Sullivan, Petty and Beatson 2011).

## RESULTS AND DISCUSSION

JF3224 isolate was initially thought to bear AsaGEI2a (Emond-Rheault et al. 2015). However, PCR assays to determine the presence of *orf21*, which is usually found in AsaGEI2a (GenBank accession number: KJ626180), indicated that JF3224 does not possess *orf21* (data not shown), suggesting that this isolate may harbor an AsaGEI2a variant. This possibility was verified by genome sequencing.

The *de novo* genome assembly of the JF3224 isolate produced 133 contigs, with an  $N_{50}$  length of 120 108 bp. The smallest and largest contigs were 599 and 382 412 bp long, respectively. The G+C content was 58.44%. The genomic alignment determined using CONTIGuator (Galardini et al. 2011) of the contigs from JF3224 with either the chromosome or the pAsa5 plasmid from the reference strain A449 (Reith et al. 2008) showed that the 75 and 12 contigs mapped on both. The BLASTn analyses (Altschul et al. 1997) of the unmapped contigs showed that contigs number 81, 76 and 73 (GenBank accession numbers: JXTA01000081, JXTA01000076 and JXTA01000073, respectively) corresponded to the high-copy-number plasmids pAsa1, pAsa2 and pAsa3, respectively (Boyd et al. 2003). Another high-copy-number plasmid (pAsa1) (Fehr et al. 2006) was also mapped on several contigs of the JF3224 genome. The remaining unmapped contigs were associated with repeated elements in the *A. salmonicida* subsp. *salmonicida* genome and corresponded to insertion sequences (IS), rRNA clusters, conserved sequences shared by prophages 1 and 2 and a 6000-bp chromosomal duplication.

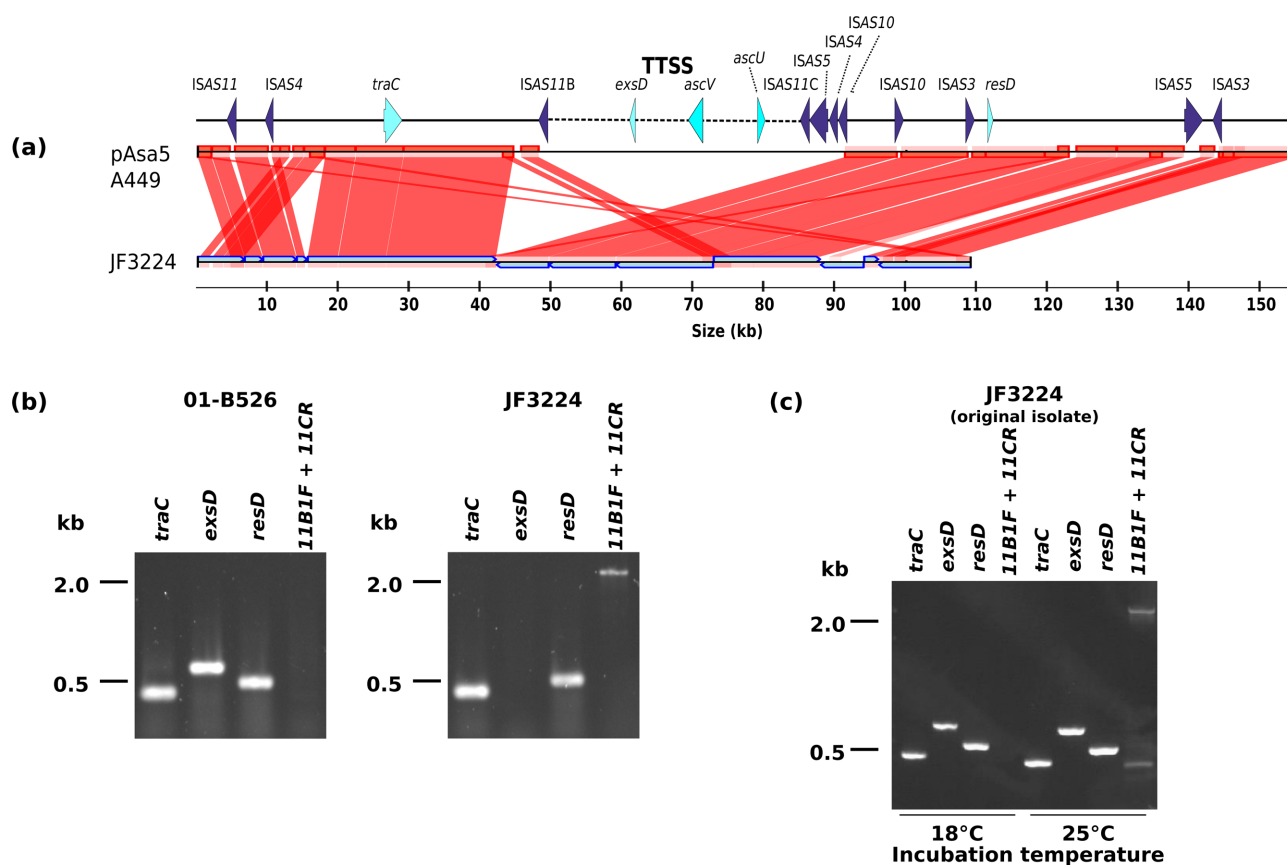
The results of the genomic alignment comparison of the reference strain (A449) and the contigs of JF3224 using CONTIGuator (Galardini et al. 2011) were visualized with ACT (Carver et al. 2005) to determine the reasons for the genome assembly breaks. The majority of the assembly breaks could be explained by repeated elements (data not shown), as reported in another study

(Vincent et al. 2014). The chromosome alignment suggested that there is a 52.8-kb sequence inserted in the JF3224 chromosome (Fig. 1a).

The detailed genome sequencing analyses indicated that JF3224 possesses a new variant of AsaGEI since it is markedly different from AsaGEI2a and other AsaGEIs (Fig. 1b). However, the novel genetic variant contained an integrase gene (*orf1*) identical to the *orf1* in AsaGEI2a (Fig. 1b). In addition, AsaGEI2a and the new genetic variant shared the same insertion site in the chromosome (data not shown). The new AsaGEI was thus called AsaGEI2b. The query cover and identities between AsaGEI2a and AsaGEI2b, which were determined using megablast (word size of 28) (Altschul et al. 1997) on the NCBI server, were 76 and 94%, respectively, indicating that approximately 25% of the genes in these AsaGEIs (i.e. AsaGEI2a and AsaGEI2b) are completely different. This level of identity is similar to that observed with AsaGEI1a and AsaGEI1b (Emond-Rheault et al. 2015).

AsaGEI2b was annotated, and the sequence is available in GenBank (accession number: KP861348). Seventy-four percent of the genes of AsaGEI2b code for hypothetical proteins. Like the other three AsaGEIs, no function has so far been attributed to this genomic element (Emond-Rheault et al. 2015). BLASTn searches (Altschul et al. 1997) using the NCBI webserver showed that AsaGEI2b is not present in any other bacterial strains.

In addition to the presence of a new AsaGEI in the JF3224 chromosome, we also observed another particularity related to the pAsa5 plasmid in the genome of this isolate. The alignment of pAsa5 from A449 (Reith et al. 2008) with the JF3224 contigs visualized with ACT (Carver et al. 2005) indicated that assembly breaks were mainly caused by repeated elements such as ISs (Fig. 2a). The assembly break sites were at the same locations as those in two other pAsa5 plasmids studied in another study (Vincent et al. 2015). The JF3224 isolate was previously



**Figure 2.** Loss of the TTSS locus from the pAsa5 plasmid of the JF3224 isolate. (a) Alignment of plasmid pAsa5 from A449 with the JF3224 contigs using CONTIGuator (Galarini et al. 2011). The locations of the 11 ISs and the *traC*, *exsD*, *ascV*, *ascU* and *resD* genes on pAsa5 were labeled using Easyfig (Sullivan et al. 2011). (b) Confirmation by PCR genotyping of the pAsa5 rearrangement in JF3224. Primers 11B1F and 11CR were used to assess B–C rearrangements while primers targeting *traC*, *exsD* and *resD* were used as controls for the presence of the various sections of pAsa5. The 01-B526 isolate was used as a control for an intact pAsa5 plasmid (Daher et al. 2011; Charette et al. 2012). (c) The original JF3224 isolate in Switzerland has an intact pAsa5 plasmid that is sensitive to insertion sequence-dependent rearrangements when incubated at 25°C. The bacteria were grown for 3 days at 18 or 25°C. Pools of approximately 20 colonies were tested in each PCR assay using the same primers as in (b).

reported to possess the *ascV* and *ascU* genes (Studer, Frey and Vanden Bergh 2013), which are located in the type three secretion system (TTSS) locus (Fig. 2a). The results of the PCR analyses (Fig. 2b) and genome sequencing performed during the present study indicated that JF3224 does not harbor the TTSS locus. We suspect that the time it took (5 days) to transport the isolate from the Swiss laboratory to the Canadian laboratory exposed the bacteria to temperatures exceeding 18°C, and that this may have caused the TTSS deletion. The instability of the TTSS locus on pAsa5, a thermolabile plasmid, has already been attributed to the incubation of *A. salmonicida* subsp. *salmonicida*, a psychrophilic bacterium, at high temperatures (25°C and higher) (Stuber et al. 2003; Daher et al. 2011; Tanaka et al. 2012). PCR assays with the appropriate primers were used to determine whether the TTSS deletion was caused by a recombination of ISAS11B and ISAS11C (Fig. 2a), as previously reported by Tanaka et al. (2012). The results indicated that ISAS11B/C were involved in the loss of the TTSS locus (Fig. 2b). This was confirmed by sequencing the amplicons (data not shown).

To verify whether pAsa5 was intact in the original isolate in Switzerland, the same PCR assays were performed on this isolate. The original isolate in Switzerland was also grown for 3 days at 25°C to determine its sensitivity to this stressful condition. As shown in Fig. 2c, the original isolate gave PCR products for the TTSS *exsD* gene, confirming the integrity of the pAsa5 plasmid.

The PCR amplification of DNA from a pool of 20 colonies grown at 25°C amplified both the *exsD* gene and the ISAS11B/C rearrangement, suggesting that some of the colonies contained a pAsa5 plasmid with an IS-dependent rearrangement while the others contained an intact plasmid. The present study provided additional proof that *A. salmonicida* subsp. *salmonicida* is sensitive to stressful conditions that can affect the genes involved in a major virulence mechanism.

The discovery of *AsaGEI2b* revealed, until proven otherwise, that *AsaGEI2a* is only found in North American isolates. Rather than illustrating a case of trade-driven intercontinental dissemination of *A. salmonicida* subsp. *salmonicida*, the draft genome of JF3224 revealed that *AsaGEIs* are more diverse than previously thought and further confirmed the usefulness of these genomic islands as indicators of the geographic origins of *A. salmonicida* subsp. *salmonicida* isolates.

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**Conflict of interest.** None declared.

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